

Package: Rabe (via r-universe)

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Title Adenine base editor analysis

Version 0.0.1

Description Base editors are emerging molecular sensors for protein-RNA interaction. This package implements a workflow for analysis of adenine base editor datasets. With minimal adjustment it can be used for systematic inquiry of any known single base-pair mutagenesis patterns. Part of the y3628 analysis suite.

License GPL (>= 3)

Encoding UTF-8

Roxygen list(markdown = TRUE)

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biocViews CellBiology, Genetics

Imports stringr, tibble, dplyr, S4Vectors, IRanges, GenomicRanges, VariantAnnotation, SummarizedExperiment

Depends R (>= 4.1)

Suggests knitr, rmarkdown

VignetteBuilder knitr

Config/pak/sysreqs make libicu-dev libpng-dev libxml2-dev libssl-dev

Repository <https://yeyuan98.r-universe.dev>

RemoteUrl <https://github.com/yeyuan98/Rabe>

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Contents

assignToGRanges	2
assignVariantIds	3
diffVariants	3
filterAdenineEditVariants	4

filterVariants	4
getConfidentVariantIds	5
getNameVarTable	6
getPrimaryVariantTable	6
getVariantTable	7
mergeVariants	7
prettyFormatSummary	8
pruneAdenineEditVariants	8
variantID2GRanges	9

Index	10
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assignToGRanges	<i>Assign variant IDs to a GRanges object based on overlap</i>
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Description

Assign variant IDs to a GRanges object based on overlap

Usage

```
assignToGRanges(variant_ids, targetGRanges, prune = TRUE)
```

Arguments

variant_ids	Character vector of variant IDs
targetGRanges	GenomicRanges::GRanges object
prune	Boolean, if TRUE will only return rows that overlap to ≥ 1 variants

Details

This function considers strandedness when finding overlap. "A/G" variants will only be assigned to "+" stranded ranges, and vice versa. However, you must make sure that only "A/G" and "T/C" variants are included in the input.

Value

GenomicRanges::GRanges with an additional mcol \$variant_ids. \$variant_ids is a IRanges::CharacterList

Examples

```
vignette("rABE-analysis")
```

assignVariantIds	<i>Assign overlapping VCF variant IDs to a GRanges</i>
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Description

Assign overlapping VCF variant IDs to a GRanges

Usage

```
assignVariantIds(range, vcf, sep = ";" )
```

Arguments

range	GenomicRanges::GRanges object
vcf	VariantAnnotation::VCF object
sep	Separator to use if >1 variants overlap to a range

Value

Input range except adding a \$variant_ids column

Examples

```
vignette("rABE-analysis")
```

diffVariants	<i>Set difference of two variant tables</i>
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Description

Set difference of two variant tables

Usage

```
diffVariants(varTableX, varTableY)
```

Arguments

varTableX	variant table X
varTableY	variant table Y

Value

variant table of X \ Y (based on \$variant_id)

Examples

```
vignette("rABE-analysis")
```

`filterAdenineEditVariants`

Filter variants to include only Adenine base editing

Description

Filter variants to include only Adenine base editing

Usage

```
filterAdenineEditVariants(assignedRange, stranded = TRUE, pruning = TRUE)
```

Arguments

- `assignedRange` GenomicRanges::GRanges with \$variant_ids column. You can assign variants with the function `assignVariantIds()`.
- `stranded` Boolean whether to consider strandedness. If TRUE, A/G editing for '+' strand, T/C for '-' strand, raising error if any strand takes '*' value.
- `pruning` Whether to prune values in \$variant_ids column. If TRUE, \$variant_ids will only retain adenine base editing events. If FALSE, \$variant_ids is not changed.

Value

GenomicRanges::GRanges containing ranges with at least one adenine base editing event.

Examples

```
vignette("rABE-analysis")
```

`filterVariants`

Filter a single merged variant table

Description

Filter a single merged variant table

Usage

```
filterVariants(mergedVarTable, perc.limits, plot = TRUE)
```

Arguments

- `mergedVarTable` Single tibble::tibble of merged variants
- `perc.limits` Lower and upper limits of median mutation percentage.
- `plot` Boolean of whether to plot a simple histogram of mutation percentages

`getConfidentVariantIds`

5

Value

Filtered merged variant table

Examples

```
vignette("rABE-analysis")
```

`getConfidentVariantIds`

Get 'confident' variants for downstream analysis

Description

Get 'confident' variants for downstream analysis

Usage

```
getConfidentVariantIds(variantTableList, print.stats = TRUE, min.occurrence = 2)
```

Arguments

`variantTableList`

A list of variant tables. Each variant table must have \$variant_id column.

`print.stats` Boolean, whether to print statistics.

`min.occurrence` Integer, minimum number of occurrences for a variant to be considered as 'confident'.

Value

A character vector of 'confident' variant IDs.

Examples

```
vignette("rABE-analysis")
```

`getNameVarTable` *Convert GRanges with assigned variants into a tibble*

Description

Convert GRanges with assigned variants into a tibble

Usage

```
getNameVarTable(assignedGRanges)
```

Arguments

`assignedGRanges`

an "assigned GRanges" (i.e., GenomicRanges::GRanges with names and `variant_ids`).

Value

A `tibble::tibble` with columns:

- `name`, character of `name(assignedGRanges)`
- `variant_ids`, list(character) of `assignedGRanges$variant_ids`

Examples

```
vignette("rABE-analysis")
```

`getPrimaryVariantTable`

Get data frame of primary variants from a VCF file

Description

Get data frame of primary variants from a VCF file

Usage

```
getPrimaryVariantTable(vcf, variant_ids)
```

Arguments

`vcf` VariantAnnotation::VCF object

`variant_ids` Identifiers to subset the VCF object

Value

A `data.frame` of counts for primary variants of each ID queried.

getVariantTable	<i>Return detailed variant information (mutation rate, counts for REF/ALT).</i>
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Description

Return detailed variant information (mutation rate, counts for REF/ALT).

Usage

```
getVariantTable(vcf, assignedRange)
```

Arguments

vcf	VariantAnnotation::VCF object
assignedRange	GenomicRanges::GRanges with \$variant_ids column. You can assign variants with the function <code>assignVariantIds()</code> .

Value

A tibble::tibble

Examples

```
vignette("rABE-analysis")
```

mergeVariants	<i>Merge a list of variant tables into one table.</i>
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Description

Merge a list of variant tables into one table.

Usage

```
mergeVariants(variantTableList, variantIDsToKeep)
```

Arguments

variantTableList	list of variant tables.
variantIDsToKeep	which variant IDs to keep in the merged frame

Value

A single tibble::tibble of merged variant tables with following columns:

- variant_id, character vector
- variant_data list of tibble::tibble, each with columns \$countREF, \$countALT, \$mutatin.perc.

Examples

```
vignette("rABE-analysis")
```

prettyFormatSummary *Format summary of a numeric vector*

Description

Format summary of a numeric vector

Usage

```
prettyFormatSummary(nums)
```

Arguments

nums	A numeric vector
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Value

A character(1) showing standard summary statistics (i.e., 0/25/50/mean/75/100 quantiles).

pruneAdenineEditVariants

Prune a GenomicRanges containing variants to retain only A->G edits

Description

Prune a GenomicRanges containing variants to retain only A->G edits

Usage

```
pruneAdenineEditVariants(rangeWithVariants, stranded = TRUE)
```

Arguments

rangeWithVariants	GenomicRanges::GRanges with \$variant_ids. Each range can contain multiple variant_ids separated by ;. Example \$variant_ids = "chr2L:1_A/G;chr2L:100_C/A"
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stranded	Boolean, whether to consider strand of the ranges
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Value

GenomicRanges::GRanges with only A->G edits

variantID2GRanges *Convert A/G variant ID vector into GenomicRanges::GRanges*

Description

Convert A/G variant ID vector into GenomicRanges::GRanges

Usage

```
variantID2GRanges(variant_ids)
```

Arguments

variant_ids Character vector of VCF variant identifier. Example: "chr2L:1_A/G"

Details

Variant type is extracted but NOT checked at all. You must make sure that only "A/G" and "T/C" are present as input.

Value

GenomicRanges::GRanges object of the variants. All widths are one as this is record of single base mutations. The following mcols are defined:

- type, "A/G" (strand = "+") or "T/C" (strand = "-") to reflect variant type.
- variant_id, the original input as record

Index

assignToGRanges, 2
assignVariantIds, 3

diffVariants, 3

filterAdenineEditVariants, 4
filterVariants, 4

getConfidentVariantIds, 5
getNameVarTable, 6
getPrimaryVariantTable, 6
getVariantTable, 7

mergeVariants, 7

prettyFormatSummary, 8
pruneAdenineEditVariants, 8

variantID2GRanges, 9